

The rationale and evidence base for a protective role of macular pigment in age-related maculopathy

E Loane,¹ C Kelliher,² S Beatty,^{1,2} J M Nolan¹

¹ Macular Pigment Research Group, Waterford Institute of Technology, Cork Road, Waterford, Ireland; ² Department of Ophthalmology, Waterford Regional Hospital, Waterford, Ireland

Correspondence to: Dr J M Nolan, Macular Pigment Research Group, Waterford Institute of Technology, Cork Road, Waterford, Ireland; jnolan@wit.ie

Accepted 11 June 2008
Published Online First XXXX

ABSTRACT

Age-related maculopathy (ARM) remains the most common cause of blind registration in people aged 50 years or over in the developed world, and its prevalence continues to rise. Although effective new treatments have become available in the recent past, these are expensive and cumbersome to the healthcare provider and to the patient, and many cases remain resistant to such therapy. There is a biologically plausible rationale whereby macular pigment, which is entirely of dietary origin, may prevent or delay the onset, or ameliorate the clinical course, of ARM. In this article, we review this rationale, and critically appraise the current evidence base germane to the use of supplements containing the macular carotenoids in patients with, or at risk of developing, ARM.

Age-related macular degeneration (AMD) is the advanced form of age-related maculopathy (ARM), and is the leading cause of blindness in people over 50 years of age in the developed world.^{1,2} The number of adults registered blind as a result of AMD in industrialised countries continues to rise, primarily due to increasing longevity.^{3,4} Beyond its inevitable impact on the individual sufferer, AMD poses a growing socio-economic challenge to modern society.⁵⁻⁷

Three dietary carotenoids, lutein (L), zeaxanthin (Z) and *meso*-zeaxanthin (*meso*-Z), accumulate at the macula, where they are collectively referred to as macular pigment (MP). L and Z are present in many foods, whereas *meso*-Z is not found in a conventional diet, although it is found in certain types of seafood.^{8,9} In recent years, the anatomic, biochemical and optical properties of MP have provoked interest in the putative protection that this pigment may confer against ARM.¹⁰

In this article, we review the literature germane to the rationale and evidence base for the putative protective effect of MP against ARM.

RATIONALE

Aetiopathogenesis of ARM

There is a consensus that genetic background and environmental/lifestyle risk factors, and an interaction between these variables, predispose to ARM. In other words, the risk that an individual's genetic make-up represents for ARM is subject to modification by environmental/lifestyle factors.¹¹ The three undisputed risk factors for ARM are: increasing age, positive family history of disease and smoking. Tobacco smoking is, therefore, the only proven environmental/lifestyle risk factor for this disease.¹²⁻¹⁴ Putative environmental/lifestyle risk factors for ARM include dietary deficiency of

antioxidants relevant to retinal health, and chronic and cumulative exposure to ambient short-wavelength (blue) light.^{15,16}

Oxidative stress

As ARM is, by definition, an age-related disorder, the free radical and the evolutionary theories of ageing are of particular relevance to its aetiopathogenesis. The free radical theory of ageing proposes that ageing and age-related disorders are the result of cumulative damage resulting from tissue reactions involving reactive oxygen intermediates (ROIs). The evolutionary theory of ageing proposes that the force of natural selection declines with increasing age, such that we may have evolved with genes, which promote morbidity and mortality once we have passed our period of procreation. In other words, genes that have a beneficial effect, or no effect, in early life are not eliminated by natural selection, even though they may have a detrimental effect in later life. Thus, both genetic background and antioxidant defences may be important for ageing and age-related morbidity, and parallels with gene-environment interactions in ARM are inescapable.

Oxidative stress occurs when the level of oxidants (ROIs) in a system exceeds the detoxifying capacity of its antioxidants.¹⁷ ROIs, which include free radicals, hydrogen peroxide and singlet oxygen, are unstable by-products of oxygen metabolism and interact with macromolecules causing damage to cells and tissues. In addition to oxygen metabolism, atmospheric pollution, asbestos exposure, tobacco use, excess consumption of alcohol, inflammation and ageing are also known to promote the production of ROIs. Interestingly, the body has an antioxidant defence system, which consists of exogenous and endogenous antioxidants, acting synergistically to quench ROIs.

The retina is ideally suited for the production of ROIs, due to its high oxygen demand, exposure to light, metabolic activities (such as retinal pigment epithelium (RPE) phagocytosis, known to generate ROIs) and its abundance of photosensitisers. Furthermore, the photoreceptor outer segments are rich in polyunsaturated fatty acids (PUFAs), such as docosahexaenoic acid (DHA), compounds that represent an ideal substrate for oxidative damage. Oxidation of PUFAs initiates a self-perpetuating chain reaction, which culminates in further production of damaging ROIs and further oxidation of neighbouring PUFAs.¹⁰

Evidence of oxidative stress can be seen in the RPE and in the neurosensory retina with increasing age, and evidence for such injury is most prominent at the macula.^{18,19} Many investigators have

Review

proposed that the age-related changes within the RPE, including an accumulation of lipofuscin within the RPE cells, represent the earliest changes that ultimately lead to ARM. Indeed, there is convincing evidence that RPE dysfunction is related, at least in part, to lipofuscin accumulation within this cell layer, which, in turn, may be attributable to incomplete phagocytosis of oxidatively damaged photoreceptor outer segments.^{20–21}

Cumulative exposure to short-wavelength light

The energy of a photon is inversely proportional to its radiation frequency; thus ultraviolet and short-wavelength (blue) light have more energy than longer wavelengths of light (eg, red and yellow). Light is composed of that part of the electromagnetic spectrum extending from ultraviolet light (with a wavelength of approximately 10–400 nm) to infrared light (with a wavelength of approximately 700–1 000 000 nm). The visible light spectrum extends from a wavelength of approximately 400–700 nm.^{22–23} The cornea and lens filter out most short-wavelength ultraviolet light. However, substantial quantities of high-energy visible light irradiate the retina, especially in young people with clear lenses. In this population, transmission of light to the retina is almost 90% at a wavelength of approximately 450 nm. In contrast, nuclear cataracts and the natural yellowing of the lens, associated with ageing, are known to limit the amount of blue light incident on the retina, with only 70–80% of incident light being transmitted below wavelengths of approximately 540 nm.²²

Since 1965, investigators have demonstrated that damage to the photoreceptors and the RPE of laboratory animals can be induced by ambient levels of visible light.²⁴ Indeed, premature induction of drusen formation in young mouse ARM models has been demonstrated with exposure to white, fluorescent light (10 000 lux, equal to outdoor sunlight) 24 h per day for 8 weeks.²⁵ However, it has been found that the blue part of the visible light spectrum is the most injurious.²³

Lipofuscin appears to be a key mediator of photo-oxidative stress, and has been shown to be a photo-inducible generator of ROIs, with the threshold for generation of these unstable molecules being lowest for light at the blue end of the visible spectrum.²⁶ Furthermore, the generation of singlet oxygen by lipofuscin increases as local oxygen tension rises, and this is particularly important in the retina, given the high oxygen tension within this tissue.²⁷ Indeed, cultures of RPE cells, laden with the lipofuscin constituent A2E, demonstrate a predilection for apoptosis when exposed to visible light, particularly the shorter wavelengths.^{28–29} In these circumstances, cellular apoptosis is secondary both to DNA damage and to mitochondrial dysfunction, caused either directly by A2E epoxides and/or ROIs or by the release of pro-apoptotic proteins (eg, cytochrome C from mitochondria).

MACULAR PIGMENT

L, Z and *meso*-Z are naturally occurring hydroxycarotenoids that accumulate at the macula (to the exclusion of other circulating carotenoids), where they are collectively known as MP.^{30–31} L and Z are not synthesised *de novo* in humans, and are entirely of dietary origin, whereas *meso*-Z is primarily formed in the retina following conversion from L.³²

An average western diet contains 1.3–3 mg/day of L and Z combined,^{33–34} with significantly more L than Z (represented by an estimated ratio of 7:1). Approximately 78% of dietary L and Z is sourced from vegetables.⁸ L is found in highest concentrations in dark green leafy vegetables, such as spinach, kale and

collard greens.⁸ Z is the major carotenoid found in corn, orange peppers and oranges, with a high mole percentage of both L and Z being found in egg yolk.⁸ Possible dietary sources of *meso*-Z include shrimp, certain marine fish and turtles, none of which is found in a typical western diet.⁹

MP represents the most conspicuous accumulation of carotenoids in the human body. Previous investigators have described the anatomic distribution of MP in the primate retina, and have demonstrated that it generally peaks at the centre of the macula, with a concentration of almost 1 mM at this location.^{31–35–36} Of note, Z is the predominant carotenoid in the foveal region, whereas L predominates in the parafoveal region.^{36–37} The concentration of *meso*-Z peaks centrally (*meso*-Z:Z ratio is 0.82 in the central retina (within 3 mm of the fovea) and 0.25 in the peripheral retina (11–21 mm from the fovea)).³⁸ The above observations are most probably attributable to the fact that retinal *meso*-Z is produced primarily by isomerisation of retinal L, thus accounting for lower relative levels of L, and higher relative levels of *meso*-Z, in the central macula, and vice versa in the peripheral macula.

Typically, the optical density of MP reaches its half-peak optical density at an average of only 1.03° (0.3 mm) retinal eccentricity.³⁹ Although MP is optically undetectable at a retinal eccentricity of 7° (2 mm), L and Z are present in the peripheral retina in minute concentrations (at distances greater than 8.7 mm from the fovea, the concentration of L and Z (combined) is about 1/300 of that within 0.25 mm of the fovea).³⁶ However, the aggregate amount of total peripheral L and Z is substantial, accounting for approximately 50% of the total amount of the carotenoids within the entire retina.^{38–40}

L, Z and *meso*-Z are intracellular compounds, separated between the cell cytosol and the cell membrane of the photoreceptor outer segment membranes, where they are possibly bound to the ubiquitous structural protein tubulin, or to specific xanthophyll-binding proteins.^{41–43} Demonstrated initially by Snodderly *et al* in primates, and more recently confirmed in human donor eyes by Trieschmann *et al*, MP has been shown to reach its maximum concentration within the photoreceptor axon layer (fibres of Henle) of the foveola, whereas outside the foveola, the highest concentrations are found both within the photoreceptor axons and within the inner and outer plexiform layers.^{35–44} It is noteworthy that MP has both a vertical (within the photoreceptor axons) and a horizontal (within the outer plexiform layer) orientation, thereby maximising prereceptor absorption of incident blue light.⁴⁵ Of note, it is believed that Z is oriented perpendicular to the cell membrane, whereas L is arranged both parallel and perpendicular to the cell membrane.⁴⁵

Functions of MP

Antioxidant

L, Z and *meso*-Z are structural isomers of one another, and their most noteworthy feature, from a biochemical perspective, rests on their high number of double bonds (and, therefore, readily available electrons).³⁰ The macular carotenoids are capable of quenching singlet oxygen, free radicals and triplet-state photosensitisers, thus limiting membrane phospholipid peroxidation.^{45–49} Kirschfeld was the first to propose the concept that carotenoids protect the macula against oxidative stress.⁵⁰ However, firm evidence that carotenoids act as antioxidants in the human retina was provided in 1997 by Khachik *et al*, who demonstrated the presence of direct oxidation products of L and Z in this tissue.⁵¹

In vitro studies of cultured human RPE cells have demonstrated enhanced survival of these cells when they are subjected to oxidative stress in the presence of Z and other antioxidant compounds, as compared with cells subjected to the same conditions in the absence of such antioxidants.⁴⁹ Of note, under in vitro conditions, L and Z are more resistant to degradation than other carotenoids when subjected to oxidative stress, an attribute which may facilitate their selective accumulation and slow biological turnover at the macula.⁵² Of the macular carotenoids, it appears that Z is a more potent antioxidant than L.^{48, 53} Studies have shown that, in conjunction with a Z-binding protein, *meso*-Z is a better antioxidant than Z; however, without the binding protein, this situation is reversed.⁴⁶

Thomson *et al* demonstrated that light-induced photoreceptor apoptosis can be limited by supplemental Z in a dose-dependent fashion in quail (the retina of which, like primates, selectively accumulates L and Z).⁵⁴ Consistent with this, Chucair *et al* have recently shown that supplemental L and Z, along with DHA, protect photoreceptors from oxidative stress-induced apoptosis, and that L and Z enhance photoreceptor differentiation.⁵⁵ In this study, in vitro cultures of retinal neurons were exposed to paraquat and hydrogen peroxide-induced oxidative stress in rats supplemented with L, Z or β -carotene (with or without DHA) and unsupplemented (control) rats. Cultures of supplemented animals exhibited less oxidative stress-induced apoptosis, and greater preservation of mitochondrial function, when compared with controls. This study provided the first evidence of direct neuroprotection of photoreceptors by the macular carotenoids. However, another recently published study by Kalariya *et al* suggests that carotenoid-derived aldehydes may actually promote oxidative stress in RPE cells.⁵⁶

Optical filter

The absorption spectrum of the macular carotenoids peaks at 460 nm, and thus MP is a filter of blue light and may limit photo-oxidative damage to retinal cells.⁵⁷ As mentioned previously, MP levels are maximal within the photoreceptor axons of the foveola and the plexiform layers of the macula.^{35, 44} Importantly, both the absorptive characteristics of MP and its location in the anterior portion of individual photoreceptors enable the pigment to attenuate the amount of blue light incident upon the photoreceptor.

It has been estimated that the quantity of visible blue light (460 nm) incident upon the photoreceptors of the macula is substantially reduced as a result of the filtering properties of MP; this reduction is estimated at approximately 40%, but varies from 3 to 100% between individuals.^{35, 58} The orientation of L (which lies both parallel and perpendicular to the cell membrane) confers greater blue-light filtering properties upon this carotenoid when compared with Z (which only lies parallel to the cell membrane), because L absorbs blue light incident from all directions.^{45, 57} However, it should be borne in mind that L, Z and *meso*-Z have slightly different absorption spectra, and thus the combination of these pigments at the macula results in the prereceptor absorption of a wider range of short-wavelengths of light than if any were present in isolation.

EVIDENCE

Scientific evidence yielded from clinical and epidemiological research may be categorised and ranked according to the perceived strength of that evidence and its freedom from bias. It is widely accepted that the strongest evidence is derived from

well-performed meta-analyses, or systematic reviews, of well-designed prospective cohort studies, and that the impact of treatment on a disease is best assessed by a comprehensive systematic review of carefully conducted randomised controlled trials (RCTs). The evidence supporting a role for MP in the prevention of ARM, or the retardation or arrestation of progression of this disease, is primarily available from observational studies and interventional (supplementation) studies.

Observational studies

Of the 10 observational studies that have examined the relationship between dietary intake of antioxidants relevant to retinal health and risk for ARM, seven have found a protective effect in association with a high intake of such antioxidants (table 1).^{16, 59–68}

Interestingly, the most recent report from the Age-Related Eye Disease Study (AREDS) found that a higher dietary intake of L and Z was independently associated with a decreased likelihood of having neovascular AMD, geographic atrophy, and large or extensive intermediate drusen.¹⁶ Furthermore, a prospective arm of the Blue Mountains Eye Study demonstrated that a high dietary intake of L and Z was associated with a reduced risk of incident neovascular AMD.⁶⁸

Recently, Chong *et al* published a meta-analysis designed to investigate the role of dietary antioxidants in the primary prevention of ARM.⁶⁹ They conducted a systematic review of seven databases, covering the years 1800–2007, and specifically directed their search to only include studies evaluating dietary intake of antioxidants relevant to retinal health in individuals without any signs of ARM at baseline. A minimum follow-up of 1 year was required for inclusion, with ARM as the primary outcome, and AMD as the secondary outcome. In total, 4192 abstracts were screened, with only 12 studies (nine prospective cohort studies and three RCTs) meeting the inclusion criteria. All studies were conducted in developed countries and reported in the last 10 years. Only five cohort studies reported on the effects of L and Z, three of which reported no association,^{62, 70, 71} one a positive (protective) association⁷² and one a negative (deleterious) association.⁶⁸ However, measurement of MP optical density was not performed in any of these studies. Furthermore, a dietary questionnaire was administered only once in each study, at baseline. The authors conceded that meta-analyses of observational data are known to have more bias than those of RCTs, and that the duration of follow-up was short in the included studies.

Of the 11 observational studies that have examined the relationship between serum levels of antioxidants and risk for ARM, seven have found an inverse (protective) association (table 2).^{59, 73–83}

However, the protective effect of MP and other retinal antioxidants, if any, rests on their ability to defend against chronic and cumulative damage and, therefore, would need to be exerted in young and middle age, and decades before disease onset. Such an analysis prompted us to investigate, in a cross-sectional fashion, MP optical density and its relationship with known and putative risk factors for ARM in 828 healthy subjects (without any evidence of retinal pathology) aged 20–60 years. In brief, we found that there was a relative lack of MP in association with the three most important risk factors for ARM (age, family history of disease and tobacco smoking). In other words, a relative lack of MP in association with risk factors for ARM, decades before disease onset, has been demonstrated in healthy subjects.⁸⁴

Review

Table 1 Observational studies examining the relationship between dietary antioxidants and risk for age-related maculopathy (ARM)

Study	Year	No. of cases	Design	Age group	Nutritional data	ARM/nutrient relationship
NHANES I	1988	3082	Cohort	45 to 74	FFQ* (vitamins A and C)	Inverse
EDCCS	1994	1994	Case-control	55 to 80	FFQ (66-item)	Inverse
BDES	1996	1968	Cohort	45 to 86	FFQ (100-item)	None
BDES	1998	1586	Population-based cohort	43 to 86	FFQ (100-item)	Inverse
BMES	1999	3654	Cross-sectional	49+	FFQ (145-item)	None
NHANES III	2001	8222	Cross-sectional	40+	FFQ (carotenoids, L and Z)	Inverse
BMES	2002	2335	Population-based cohort	49+	FFQ (145-item)	None
NHS and HPFS	2004	118 428	Prospective follow-up	50+	FFQ (vitamins and carotenoids)	Inverse
AREDS	2007	4513	Case-control	55 to 80	FFQ (carotenoids, L and Z)	Inverse
BMES	2007	2454	Population-based cohort	49+	FFQ (145-item, L and Z)	Inverse

AREDS, Age-Related Eye Disease Study; BDES, XXXX; BMES, XXXX; EDCCS, XXXX; FFQ, XXXX; HPFS, XXXX; L, lutein; NHANES, XXXX; NHS, XXXX; Z, zeaxanthin.

Interventional studies

Koh *et al* reported that serum and macular carotenoid response in patients with ARM is similar to that of control subjects.⁸⁵ Richer *et al* have shown that ARM patients with the lowest MP optical density at baseline exhibit the greatest augmentation of MP optical density following supplementation with macular carotenoids.⁸⁶ Indeed, this finding is consistent with the recently published Lutein Nutrition effects measured by Autofluorescence (LUNA) study, where subjects with low-baseline MP optical density were more likely to exhibit a dramatic rise in MP optical density, or to exhibit no rise in MP optical density, in response to supplements than were subjects with medium- to high-baseline MP optical density values.⁸⁷ This latter finding suggests that low MP optical density values are attributable to a dietary lack of the macular carotenoids (where supplementation causes a dramatic rise in MP optical density) or to an inability of the retina to accumulate or stabilise these carotenoids (subjects who did not exhibit augmentation of MP optical density following supplementation, in spite of expected and observed increases in serum carotenoid concentrations).

Studies investigating supplemental β -carotene and vitamin E have failed to identify a beneficial effect of such supplements on the incidence and/or progression of ARM.^{88–89} The AREDS investigated the use of a high-dose antioxidant formulation (vitamin C, vitamin E, β -carotene and zinc) on the progression of ARM in a 5-year prospective RCT.⁹⁰ Over 4700 patients, aged 55–80 years, were enrolled in this study, which demonstrated that patients with moderate to advanced ARM (extensive intermediate size drusen, at least one large drusen, non-central geographic atrophy in one/both eyes, or vision loss due to ARM in one eye) exhibited a 25% risk reduction in progression to advanced ARM when supplemented with zinc plus antioxi-

dants. Unfortunately, however, the AREDS formulation did not contain the macular carotenoids (L, Z or *meso*-Z), as they were not commercially available at the inception of that study.

The first investigation of an L-fortified diet (consisting of dark green leafy vegetables and spinach) or supplementation with L-based formulations, in ARM patients, was conducted in 1999 by Richer.⁹¹ This small pilot study, with short follow-up, reported a beneficial effect on visual function in one or both eyes of patients, and led the way for a formal RCT evaluation of L-based supplements in ARM.

In 2004, the Lutein Antioxidant Supplementation Trial (LAST) was undertaken to investigate whether nutritional supplementation with L alone, or L together with antioxidants, vitamins and minerals, improved or stabilised visual function in patients with advanced atrophic ARM.⁹² This study was a prospective, 12-month, double-masked RCT involving 90 patients with ARM. The investigators reported that visual function improved with L supplementation alone, or with L supplementation in combination with other antioxidants, when compared with control subjects.⁸⁶ However, it should be noted that none of the above studies was designed to investigate whether antioxidant supplements have any effect on the primary prevention of ARM, as subjects without disease were not enrolled in these studies.

The AREDS II trial, which is currently in progress, is a placebo-controlled RCT involving 4,000 subjects with moderate to advanced ARM that is investigating the effects of supplementation with high doses of the macular carotenoids and omega-3 PUFAs, in addition to the original AREDS formulation (with the exception of β -carotene). Unfortunately, however, measurement of MP levels is not forming part of the investigation of AREDS II (with the exception of a few study

Table 2 Observational studies examining the relationship between serum antioxidants and risk for age-related maculopathy (ARM)

Study	Year	No. of cases/ no. of controls	Design	Age group	Nutritional data	ARM/serum antioxidant relationship
Blumenkranz <i>et al</i>	1986	26/23	Case-control	–	Vitamins A, C and E	None
Tsang <i>et al</i>	1992	80/86	Case-control	–	Vitamin E and selenium	None
EDCCS	1992	421/615	Case-control	–	Carotenoids	Inverse
EDCCS	1993	421/615	Case-control	55 to 80	Vitamins C, E carotenoids and zinc	Inverse
BLSA	1994	870	Cohort	40+	Vitamins, retinol and β -carotene	Inverse
BDES	1995	167	Case-control	43 to 86	Vitamin E and carotenoids	None
BMES	1997	156/156	Case-control	–	Vitamin E and β -carotene	None
Belda <i>et al</i>	1999	25/15	Case-control	60+	Vitamin E and zinc	Inverse
POLA	1999	2584	Cross-sectional	–	Vitamin E	Inverse
NHANES III	2001	8222	Cross-sectional	40+	L and Z (combined)	Inverse
Gale <i>et al</i>	2003	380	Cross-sectional	66 to 75	L and Z (separate)	Inverse (Z only)

BDES, XXXX; BLSA, XXXX; BMES, XXXX; EDCCS, XXXX; L, lutein; NHANES, XXXX; POLA, XXXX; Z, zeaxanthin.

sites). Another study that is currently in progress is the Carotenoids in Age-Related Maculopathy (CARM) study, which is a placebo-controlled, double-masked RCT, designed to investigate the potential benefits of supplementation with L, Z and coantioxidants on the progression of ARM.⁹³

COMMENT

Although the notion that MP protects against ARM remains a hypothesis, the rationale in support of this view is biologically plausible and supported by the findings of in vitro and animal studies, in which L and/or Z have been shown to protect photoreceptors against oxidative injury.

We understand that ophthalmologists currently find themselves in a difficult position when attempting to make sound and evidence-based recommendations to patients with ARM. It is true that the AREDS formulation remains the only formulation that has been shown, in the context of a well-designed RCT, to be of benefit in ARM. However, the AREDS formulation contains β -carotene, which is associated with an increased risk of lung cancer among smokers.⁹⁴ Also, the doses of vitamin C, vitamin E and zinc in the AREDS formulation far exceed the European Union (EU) upper safety limits.⁹⁵ However, it is difficult to ignore the basic implication of the AREDS, namely that antioxidants are beneficial for patients with ARM. It is such an interpretation that has encouraged the nutraceutical industry to promote the use of antioxidant supplements that do not include β -carotene, are EU-compliant and contain the macular carotenoids. One may understand why an ophthalmologist, in the absence of an evidence base but in the presence of a biologically plausible rationale, might recommend such a supplement in view of the lack of other available putative or proven preventive measures against ARM. The patient, who may have already lost vision in one eye, often explains how they wish to participate actively in risk reduction against further visual loss, and how they are unwilling to wait for a conclusive evidence base. Nevertheless, under these circumstances, it is incumbent upon the ophthalmologist to inform patients with ARM that such supplements have not been proven to protect against development, or progression, of ARM.

In conclusion, we await the outcomes of several RCTs before a meaningful comment can be made upon the potential beneficial effects of supplemental L and Z in patients with ARM. However, the benefits of L and Z, if any, relate to the ability of these compounds to protect against chronic and cumulative damage, and therefore MP may indeed be important in preventing and delaying the onset of ARM. The ongoing RCTs are not designed to test this hypothesis, which would require longitudinal data involving serial MP measurements in a large number of subjects over a period of at least 20 years.

Funding: EL is currently conducting research towards a PhD degree, part-funded by a grant from Bausch & Lomb Ireland Limited.

Competing interests: CK and EL: None. SB and JMN do consultancy work for nutraceutical companies, in a personal capacity, and as directors of NutraSight Consultancy Limited.

REFERENCES

- Bressler NM.** Age-related macular degeneration is the leading cause of blindness. *JAMA* 2004;**291**:1900–1.
- Congdon NG, Friedman DS, Lietman T.** Important causes of visual impairment in the world today. *JAMA* 2003;**290**:2057–60.
- Bunce C, Wormald R.** Causes of blind certifications in England and Wales: April 1999–March 2000. *Eye* 2007.
- Kelliher C, Kenny D, O'Brien C.** Trends in blind registration in the adult population of the Republic of Ireland 1996–2003. *Br J Ophthalmol* 2006;**90**:367–71.
- van LR, Klaver CC, Vingerling JR, et al.** Epidemiology of age-related maculopathy: a review. *Eur J Epidemiol* 2003;**18**:845–54.
- Augustin A, Sahel JA, Bandello F, et al.** Anxiety and depression prevalence rates in age-related macular degeneration. *Invest Ophthalmol Vis Sci* 2007;**48**:1498–503.
- Bandello F, Lafuma A, Berdeaux G.** Public health impact of neovascular age-related macular degeneration treatments extrapolated from visual acuity. *Invest Ophthalmol Vis Sci* 2007;**48**:96–103.
- Sommerburg O, Keunen JEE, Bird AC, et al.** Fruits and vegetables that are sources for lutein and zeaxanthin: the macular pigment in human eyes. *Br J Ophthalmol* 1998;**82**:907–10.
- Maoka T, Arai A, Shimizu M, et al.** The first isolation of enantiomeric and meso-zeaxanthin in nature. *Comp Biochem Physiol B* 1986;**83**:121–4.
- Beatty S, Koh HH, Henson D, et al.** The role of oxidative stress in the pathogenesis of age-related macular degeneration. *Surv Ophthalmol* 2000;**45**:115–34.
- Schork NJ.** Genetics of complex disease: approaches, problems, and solutions. *Am J Respir Crit Care Med* 1997;**156**:103–9S.
- Klein R, Klein BEK, Moss SE.** Relation of smoking to the incidence of age-related maculopathy—The Beaver Dam Eye Study. *Am J Epidemiol* 1998;**147**:103–10.
- Tan JS, Mitchell P, Kifley A, et al.** Smoking and the long-term incidence of age-related macular degeneration: the Blue Mountains Eye Study. *Arch Ophthalmol* 2007;**125**:1089–95.
- Chakravarthy U, Augood C, Bentham GC, et al.** Cigarette smoking and age-related macular degeneration in the EUREYE Study. *Ophthalmology* 2007;**114**:1157–63.
- Tomany SC, Cruickshanks KJ, Klein R, et al.** Sunlight and the 10-year incidence of age-related maculopathy—The Beaver Dam eye study. *Arch Ophthalmol* 2004;**122**:750–7.
- SanGiovanni JP, Chew EY, Clemons TE, et al.** The relationship of dietary carotenoid and vitamin A, E, and C intake with age-related macular degeneration in a case-control study: AREDS Report No. 22. *Arch Ophthalmol* 2007;**125**:1225–32.
- McCord JM.** The evolution of free radicals and oxidative stress. *Am J Med* 2000;**108**:652–9.
- Suzuki M, Kamei M, Itabe H, et al.** Oxidized phospholipids in the macula increase with age and in eyes with age-related macular degeneration. *Mol Vis* 2007;**13**:772–8.
- Lu L, Hackett SF, Mincey A, et al.** Effects of different types of oxidative stress in RPE cells. *J Cell Physiol* 2006;**206**:119–25.
- Sundelin S, Wihlmark U, Nilsson SE, et al.** Lipofuscin accumulation in cultured retinal pigment epithelial cells reduces their phagocytic capacity. *Curr Eye Res* 1998;**17**:851–7.
- Kennedy CJ, Rakoczy PE, Constable IJ.** Lipofuscin of the retinal pigment epithelium: a review. *Eye* 1995;**9**:763–71.
- Algvere PV, Marshall J, Seregard S.** Age-related maculopathy and the impact of blue light hazard. *Acta Ophthalmol Scand* 2006;**84**:4–15.
- Wu J, Seregard S, Algvere PV.** Photochemical damage of the retina. *Surv Ophthalmol* 2006;**51**:461–81.
- Noell WK, Walker VS, Kang BS, et al.** Retinal damage by light in rats. *Invest Ophthalmol* 1966;**5**:450–73.
- Imamura Y, Noda S, Hashizume K, et al.** Drusen, choroidal neovascularization, and retinal pigment epithelium dysfunction in SOD1-deficient mice: a model of age-related macular degeneration. *Proc Natl Acad Sci U S A* 2006;**103**:11282–7.
- Boulton M, Dontsov A, Jarvisvans J, et al.** Lipofuscin is a photoinducible free-radical generator. *J Photochem Photobiol B-Biol* 1993;**19**:201–4.
- Rozanowska M, Wessels J, Boulton M, et al.** Blue light-induced singlet oxygen generation by retinal lipofuscin in non-polar media. *Free Rad Biol Med* 1998;**24**:1107–12.
- Shaban H, Borras C, Vina J, et al.** Phosphatidylglycerol potently protects human retinal pigment epithelial cells against apoptosis induced by A2E, a compound suspected to cause age-related macula degeneration. *Exp Eye Res* 2002;**75**:99–108.
- Sparrow JR, Vollmer-Snarr HR, Zhou J, et al.** A2E-epoxides damage DNA in retinal pigment epithelial cells. Vitamin E and other antioxidants inhibit A2E-epoxide formation. *J Biol Chem* 2003;**278**:18207–13.
- Bone RA, Landrum JT, Hime GW, et al.** Stereochemistry of the human macular carotenoids. *Invest Ophthalmol Vis Sci* 1993;**34**:2033–40.
- Bone RA, Landrum JT, Tarsis SL.** Preliminary identification of the human macular pigment. *Vision Res* 1985;**25**:1531–5.
- Neuringer M, Sandstrom MM, Johnson EJ, et al.** Nutritional manipulation of primate retinas. I: effects of lutein or zeaxanthin supplements on serum and macular pigment in xanthophyll-free rhesus monkeys. *Invest Ophthalmol Vis Sci* 2004;**45**:3234–43.
- Nebeling LC, Forman MR, Graubard BI, et al.** Changes in carotenoid intake in the United States: The 1987 and 1992 National Health Interview Surveys. *J Am Diet Assoc* 1997;**97**:991–6.
- Nebeling LC, Forman MR, Graubard BI, et al.** The impact of lifestyle characteristics on carotenoid intake in the United States: The 1987 National Health Interview Survey. *Am J Publ Health* 1997;**87**:268–71.
- Snodderly DM, Auran JD, Delori FC.** The macular pigment. II. Spatial distribution in primate retinas. *Invest Ophthalmol Vis Sci* 1984;**25**:674–85.
- Bone RA, Landrum JT, Fernandez L, et al.** Analysis of the macular pigment by HPLC—Retinal distribution and age study. *Invest Ophthalmol Vis Sci* 1988;**29**:843–9.
- Snodderly DM, Handelman GJ, Adler AJ.** Distribution of individual macular pigment carotenoids in central retina of macaque and squirrel monkeys. *Invest Ophthalmol Vis Sci* 1991;**32**:268–79.
- Bone RA, Landrum JT, Friedes LM, et al.** Distribution of lutein and zeaxanthin stereoisomers in the human retina. *Exp Eye Res* 1997;**64**:211–18.

Review

39. **Hammond BR**, Wooten BR, Snodderly DM. Individual variations in the spatial profile of human macular pigment. *J Opt Soc Am A Optics Image Sci Vis* 1997;**14**:1187–96.
40. **Sommerburg O**, Siems WG, van Kuijk FJ. Localization of carotenoids in different eye tissues. *Biofactors* 2000;**11**:3–6.
41. **Bernstein PS**, Balashov NA, Tsong ED, *et al*. Retinal tubulin binds macular carotenoids. *Invest Ophthalmol Vis Sci* 1997;**38**:167–75.
42. **Rapp LM**, Maple SS, Choi JH. Lutein and zeaxanthin concentrations in rod outer segment membranes from perifoveal and peripheral human retina. *Invest Ophthalmol Vis Sci* 2000;**41**:1200–9.
43. **Bhosale P**, Larson AJ, Frederick JM, *et al*. Identification and characterization of a Pi isoform of glutathione S-transferase (GSTP1) as a zeaxanthin-binding protein in the macula of the human eye. *J Biol Chem* 2004;**279**:49447–54.
44. **Trieschmann M**, van Kuijk FJ, Alexander R, *et al*. Macular pigment in the human retina: histological evaluation of localization and distribution. *Eye* 2007.
45. **Sujak A**, Gabrielska J, Grudzinski W, *et al*. Lutein and zeaxanthin as protectors of lipid membranes against oxidative damage: The structural aspects. *Arch Biochem Biophys* 1999;**371**:301–7.
46. **Bhosale P**, Bernstein PS. Synergistic effects of zeaxanthin and its binding protein in the prevention of lipid membrane oxidation. *Biochim Biophys Acta* 2005;**1740**:116–21.
47. **Trevithick-Sutton CC**, Foote CS, Collins M, *et al*. The retinal carotenoids zeaxanthin and lutein scavenge superoxide and hydroxyl radicals: a chemiluminescence and ESR study. *Mol Vis* 2006;**12**:1127–35.
48. **Kim SR**, Nakanishi K, Itagaki Y, *et al*. Photooxidation of A2-PE, a photoreceptor outer segment fluorophore, and protection by lutein and zeaxanthin. *Exp Eye Res* 2006;**82**:828–39.
49. **Wrona M**, Rozanowska M, Sarna T. Zeaxanthin in combination with ascorbic acid or alpha-tocopherol protects ARPE-19 cells against photosensitized peroxidation of lipids. *Free Rad Biol Med* 2004;**36**:1094–1101.
50. **Kirschfeld K**. Carotenoid pigments: their possible role in protecting against photooxidation in eyes and photoreceptor cells. *Proc R Soc Lond B Biol Sci* 1982;**216**:71–85.
51. **Khachik F**, Bernstein PS, Garland DL. Identification of lutein and zeaxanthin oxidation products in human and monkey retinas. *Invest Ophthalmol Vis Sci* 1997;**38**:1802–11.
52. **Siems WG**, Sommerburg O, van Kuijk FJ. Lycopene and beta-carotene decompose more rapidly than lutein and zeaxanthin upon exposure to various pro-oxidants in vitro. *Biofactors* 1999;**10**:105–13.
53. **Cantrell A**, McGarvey DJ, Truscott TG, *et al*. Singlet oxygen quenching by dietary carotenoids in a model membrane environment. *Arch Biochem Biophys* 2003;**412**:47–54.
54. **Thomson LR**, Toyoda Y, Langner A, *et al*. Elevated retinal zeaxanthin and prevention of light-induced photoreceptor cell death in quail. *Invest Ophthalmol Vis Sci* 2002;**43**:3538–49.
55. **Chucacir AJ**, Rotstein NP, SanGiovanni JP, *et al*. Lutein and zeaxanthin protect photoreceptors from apoptosis induced by oxidative stress: relation with docosahexaenoic acid. *Invest Ophthalmol Vis Sci* 2007;**48**:5168–77.
56. **Kalariya NM**, Ramana KV, Srivastava SK, *et al*. Carotenoid derived aldehydes-induced oxidative stress causes apoptotic cell death in human retinal pigment epithelial cells. *Exp Eye Res* 2008;**86**:70–80.
57. **Junghans A**, Sies H, Stahl W. Macular pigments lutein and zeaxanthin as blue light filters studied in liposomes. *Arch Biochem Biophys* 2001;**391**:160–4.
58. **Stringham JM**, Hammond BR, Wooten BR, *et al*. Compensation for light loss resulting from filtering by macular pigment: relation to the S-cone pathway. *Optom Vis Sci* 2006;**83**:887–94.
59. **Mares-Perlman JA**, Fisher AI, Klein R, *et al*. Lutein and zeaxanthin in the diet and serum and their relation to age-related maculopathy in the Third National Health and Nutrition Examination Survey. *Am J Epidemiol* 2001;**153**:424–32.
60. **Mares-Perlman JA**, Klein R, Klein BEK, *et al*. Association of zinc and antioxidant nutrients with age-related maculopathy. *Arch Ophthalmol* 1996;**114**:991–7.
61. **Seddon JM**, Ajani UA, Sperduto RD. Dietary carotenoids, vitamins A, C, and E, and advanced age-related macular degeneration. *JAMA* 1994;**272**:1413–20.
62. **VandenLangenberg GM**, Mares-Perlman JA, Klein R, *et al*. Associations between antioxidant and zinc intake and the 5-year incidence of early age-related maculopathy in the Beaver Dam Eye Study. *Am J Epidemiol* 1998;**148**:204–14.
63. **Smith W**, Mitchell P, Webb K, *et al*. Dietary antioxidants and age-related maculopathy—The Blue Mountains Eye Study. *Ophthalmology* 1999;**106**:761–7.
64. **Flood V**, Wang JJ, Manzi F, *et al*. Dietary antioxidant intake and incidence of early age-related maculopathy: the Blue Mountains Eye Study. *Ophthalmology* 2002;**109**:2272–8.
65. **Cho EY**, Seddon JM, Rosner B, *et al*. Prospective study of intake of fruits, vegetables, vitamins, and carotenoids and risk of age-related maculopathy. *Arch Ophthalmol* 2004;**122**:883–92.
66. **Goldberg J**, Flowerdrew G, Smith E. Factors associated with age-related macular degeneration—analysis of data from NHANES I. *Am J Epidemiol* 1998;**128**:700–11.
67. **Snellen ELM**, Verbeek ALM, van den Hoogen GWP, *et al*. Neovascular age-related macular degeneration and its relationship to antioxidant intake. *Acta Ophthalmol Scand* 2002;**80**:368–71.
68. **Tan JS**, Wang JJ, Flood V, *et al*. Dietary antioxidants and the long-term incidence of age-related macular degeneration. The Blue Mountains Eye Study. *Ophthalmology* **115**:334–41. 4
69. **Chong EW**, Wong TY, Kreis AJ, *et al*. Dietary antioxidants and primary prevention of age related macular degeneration: systematic review and meta-analysis. *BMJ* 2007;**335**:755.
70. **van LR**, Boekhoorn S, Vingerling JR, *et al*. Dietary intake of antioxidants and risk of age-related macular degeneration. *JAMA* 2005;**294**:3101–7.
71. **Moeller SM**, Parekh N, Tinker L, *et al*. Associations between intermediate age-related macular degeneration and lutein and zeaxanthin in the Carotenoids in Age-related Eye Disease Study (CAREDS): ancillary study of the Women's Health Initiative. *Arch Ophthalmol* 2006;**124**:1151–62.
72. **Cho E**, Seddon JM, Rosner B, *et al*. Prospective study of intake of fruits, vegetables, vitamins, and carotenoids and risk of age-related maculopathy. *Arch Ophthalmol* 2004;**122**:883–92.
73. **Blumenkranz MS**, Russell SR, Robey MG, *et al*. Risk-factors in age-related maculopathy complicated by choroidal neovascularization. *Ophthalmology* 1986;**93**:552–8.
74. **Yannuzzi LA**, Sorenson JA, Sobel RS, *et al*. Risk-factors for neovascular age-related macular degeneration. *Arch Ophthalmol* 1992;**110**:1701–8.
75. **Gale CR**, Hall NF, Phillips DIW, *et al*. Lutein and zeaxanthin status and risk of age-related macular degeneration. *Invest Ophthalmol Vis Sci* 2003;**44**:2461–5.
76. **Sperduto RD**. Antioxidant status and neovascular age-related macular degeneration. *Arch Ophthalmol* 1993;**111**:104–9.
77. **Mares-Perlman JA**, Brady WE, Klein R, *et al*. Serum antioxidants and age-related macular degeneration in a population based case control study. *Arch Ophthalmol* 1995;**113**:1518–23.
78. **Delcourt C**, Cristol JP, Tessier F, *et al*. Age-related macular degeneration and antioxidant status in the POLA study. *Arch Ophthalmol* 1999;**117**:1384–90.
79. **Smith W**, Mitchell P, Rochester C. Serum beta carotene, alpha tocopherol, and age-related maculopathy: the Blue Mountains Eye Study. *Am J Ophthalmol* 1997;**124**:838–40.
80. **Aoki K**, Ito Y, Sasaki R, *et al*. Smoking, alcohol drinking and serum carotenoids levels. *Japan J Cancer Res* 1987;**78**:1049–56.
81. **West S**, Vitale S, Hallfrisch J, *et al*. Are antioxidants or supplements protective for age-related macular degeneration? *Arch Ophthalmol* 1994;**112**:222–7.
82. **Belda JI**, Roma J, Vilela C, *et al*. Serum vitamin E levels negatively correlate with severity of age-related macular degeneration. *Mech Ageing Dev* 1999;**107**:159–64.
83. **Tsang NC**, Penfold PL, Snitch PJ, *et al*. Serum levels of antioxidants and age-related macular degeneration. *Doc Ophthalmol* 1992;**81**:387–400.
84. **Nolan JM**, Stack J, O' Donovan O, *et al*. Risk factors for age-related maculopathy are associated with a relative lack of macular pigment. *Exp Eye Res* 2007;**84**:61–74.
85. **Koh HH**, Murray IJ, Nolan D, *et al*. Serum and macular responses to lutein supplement in subjects with and without age-related maculopathy: a pilot study. *Exp Eye Research* 2004;**79**:21–7.
86. **Richer S**, Devenport J, Lang JC. LAST II: Differential temporal responses of macular pigment optical density in patients with atrophic age-related macular degeneration to dietary supplementation with xanthophylls. *Optometry* 2007;**78**:213–19.
87. **Trieschmann M**, Beatty S, Nolan JM, *et al*. Changes in macular pigment optical density and serum concentrations of its constituent carotenoids following supplemental lutein and zeaxanthin: the LUNA study. *Exp Eye Res* 2007;**84**:718–28.
88. **Taylor HR**, Tikellis G, Robman LD, *et al*. Vitamin E supplementation and macular degeneration: randomised controlled trial. *Br Med J* 2002;**325**:11–14.
89. **Teikari JM**, Laatikainen L, Virtamo J, *et al*. Six-year supplementation with alpha-tocopherol and beta-carotene and age-related maculopathy. *Acta Ophthalmol Scand* 1998;**76**:224–9.
90. **Kasoff A**, Kasoff J, Buehler J, *et al*. A randomized, placebo-controlled, clinical trial of high-dose supplementation with vitamins C and E, beta carotene, and zinc for age-related macular degeneration and vision loss—AREDS Report No. 8. *Arch Ophthalmol* 2001;**119**:1417–36.
91. **Richer S**. ARMD—pilot (case series) environmental intervention data. *J Am Optom Assoc* 1999;**70**:24–36.
92. **Richer S**, Stiles W, Statkute L, *et al*. Double-masked, placebo-controlled, randomized trial of lutein and antioxidant supplementation in the intervention of atrophic age-related macular degeneration: the Veterans LAST study (Lutein Antioxidant Supplementation Trial). *Optometry* 2004;**75**:216–30.
93. **Neelam K**, Hogg RE, Stevenson M, *et al*. Carotenoids in age-related maculopathy: Design and methods. *Ophthalmic Epidemiol*. In press.
94. **Albanes D**, Heinonen OP, Taylor PR, *et al*. Alpha-Tocopherol and beta-carotene supplements and lung cancer incidence in the alpha-tocopherol, beta-carotene cancer prevention study: effects of base-line characteristics and study compliance. *J Natl Cancer Inst* 1996;**88**:1560–70.
95. **Anon**. Vitamins: what they do and where to find them (EU Food Information Council). 2008. 6

Authors QueriesJournal: **British Journal of Ophthalmology**Paper: **bj135566**Title: **The rationale and evidence base for a protective role of macular pigment in age-related maculopathy**

Dear Author

During the preparation of your manuscript for publication, the questions listed below have arisen. Please attend to these matters and return this form with your proof. Many thanks for your assistance

Query Reference	Query	Remarks
1	You have opted not to pay a fee to make your article free online (Unlocked). If you wish to change your mind, please indicate this clearly and provide invoice address and contact details when you return your proofs. This is the last point in the production process where you can choose to Unlock your paper; the cost of this service is £1,200/\$2,220/€1,775 (+VAT) and further details can be found at http://bjo.bmj.com/info/unlocked.dtl	
2	Please define the abbreviations in table 1 (in the footnote). Also, what does the asterisk denote?	
3	Please define the abbreviations in table 2 (in the footnote).	
4	I have updated reference 68. Please check	
5	Please update "in press" reference, if details are known.	
6	Reference 95: If this is an internet communication, please provide full details of the URL and author/s or correspondents	